

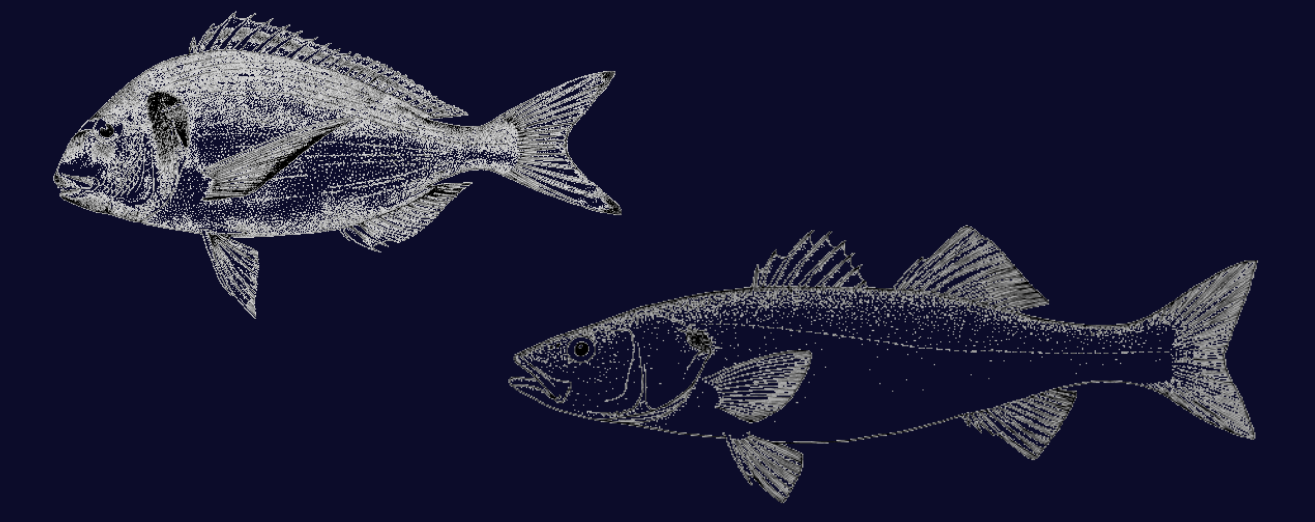
COMPARATIVE PARASITOLOGICAL STUDIES ON CULTURED GILTHEAD SEABREAM (*SPARUS AURATA*) AND EUROPEAN SEABASS (*DICENTRARCHUS LABRAX*) IN PORTUGUESE FISH FARMS

P. Ramos^{*,†}, I. Pinto^{*} and M.M. Oliveira^{*,¶}

^{*}Portuguese Institute for the Ocean and Atmosphere, IPMA, I.P., Av. Doutor Alfredo Magalhães Ramalho, 6, 1495-165 Algés, PT

[†]CIIMAR, Interdisciplinary Centre of Marine and Environmental Research, University of Porto, Rua das Bragas 289, 4050-123 Porto, PT

[¶]Lusófona University, ECEO, Campo Grande 376, 1749-024 Lisboa, PT



INTRODUCTION : Portugal is a country surrounded by the sea and thus with a great aquaculture potential. The gilthead seabream (*Sparus aurata*) and European seabass (*Dicentrarchus labrax*) are two of the most important fish commonly produced in earth ponds, where the water is renewed with the tides. The gradual increase of production of fish resulted in parasitological outbreaks in this production system with significant effects on commercial production. Thus, focus has been placed on parasitic diseases in enterprises.

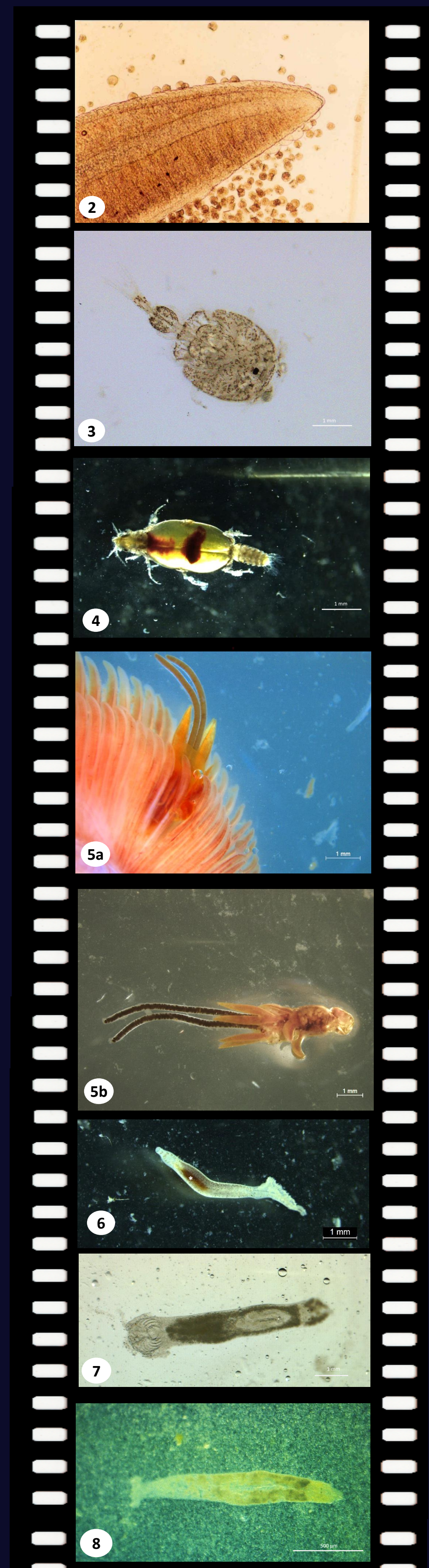
MATERIAL AND METHODS: Parasitological examinations were conducted on 271 fishes (138 seabass and 133 gilthead seabream) from semi-intensive fish farms from north, centre and south of Portugal in 2014 and 2023. Fish farms 1, 2 and 7 are supplied with brackish water by Sado estuary - Centre. Fish farms (3 and 8) and (4 and 9) are supplied with sea water from Ria de Alvor Lagoon and Ria Formosa respectively - South. Fish farms (5 and 6) and 10 are supplied with sea water from Ria de Aveiro Lagoon – North (Fig. 1).

The body surface (skin, fins) and the gills of the fish were examined for ectoparasites. Each gill arch was examined separately. The visceral cavity was opened and internal organs (external surfaces) scanned to nematodes. Biometrical measures were recorded. Prevalence, abundance and mean intensity of the infection were calculated (table 1 and 2.). Descriptive statistics were performed for each sampling site and compared the parasitic infection rates of gilthead seabream and European seabass from semi-intensive Portuguese fish farms in 2014 and 2023.

RESULTS

2014										
Sampling site	Sampling date	Species	n	Weight (g)	Length (cm)	Found parasites	Parasites n=	Prevalence (%)	Abundance (± DP)	Intensity (± DP)
1 (Sado river)	12.02.2014	<i>Sparus aurata</i>	7	156.44 (±38.83)	21.52 (±1.69)	<i>Caligus</i> sp.	1	14	0.14 (±0.38)	1
		<i>Dicentrarchus labrax</i>	3	162.72 (±22.15)	26.2 (±1.06)	<i>Caligus</i> sp.	3	33	1 (±1.73)	3
						<i>Lernanthropus kroyeri</i>	3	33	1 (±1.73)	3
2 (Sado river)	12.02.2014	<i>Sparus aurata</i>	10	441.83 (±61.69)	27.85 (±5.18)	<i>Diplectanum aequans</i>	449	83	37.4 (±44.1)	44.9 (±44.8)
3 (Ria de Alvor Lagoon)	10.03.2014	<i>Dicentrarchus labrax</i>	12	314.73 (±39.74)	26.36 (±1.35)	<i>Trichodina</i> sp.	-	8	-	-
						<i>Diplectanum aequans</i>	187	100	18.7 (±10.9)	18.7 (±10.9)
4 (Ria Formosa lagoon)	10.03.2014	<i>Dicentrarchus labrax</i>	10	342.01 (±42.82)	30.44 (±0.97)	<i>Diplectanum aequans</i>	329	92	25.3 (±24.1)	27.4 (±23.8)
						<i>Trichodina</i> sp.	-	16	-	-
5 (Ria de Aveiro)	20.05.2014	<i>Dicentrarchus labrax</i>	13	94.73 (±18.83)	20.35 (±1.41)	<i>Diplectanum aequans</i>	127	100	12.7 (±6.0)	12.7 (±6.0)
						<i>Trichodina</i> sp.	-	10	-	-
6 (Ria de Aveiro)	20.05.2014	<i>Dicentrarchus labrax</i>	10	300.42 (±81.27)	35.1 (±19.63)	<i>Praniza</i>	5	40	0.5 (±0.73)	1.25 (±0.5)
						<i>Sparocotyle chrysophrii</i>	14	67	1.56 (±1.51)	2.33 (±1.21)
5 (Ria de Aveiro)	28.10.2014	<i>Dicentrarchus labrax</i>	16	210.90 (±63.63)	27.03 (±2.48)	<i>Sparocotyle chrysophrii</i>	1	11	0.11 (±0.33)	1
						<i>Diplectanum aequans</i>	448	100	28 (±21.1)	28 (±21.1)
6 (Ria de Aveiro)	28.10.2014	<i>Dicentrarchus labrax</i>	15	339.83 (±85.20)	31.33 (±3.30)	<i>Diplectanum aequans</i>	532	100	35.47 (±22.8)	35.47 (±22.8)
						<i>Diplectanum aequans</i>	133	64	11.7 (±14.9)	17.6 (±15.5)
1 (Sado river)	9.12.2014	<i>Dicentrarchus labrax</i>	11	229.55 (±46.36)	27.45 (±1.53)	<i>Praniza</i>	1	9	0.09 (±0.30)	1
						<i>Dactylogyrus</i> sp.	77	100	7.7 (±2.87)	7.7 (±2.87)
2 (Sado river)	9.12.2014	<i>Sparus aurata</i>	10	311.37 (±23.43)	26.36 (±0.55)	<i>Dactylogyrus</i> sp.	2	50	1 (±1.41)	2
						<i>Dactylogyrus</i> sp.	96	100	9.6 (±3.1)	9.6 (±3.1)
					<i>Sparocotyle chrysophrii</i>	7	40	0.7 (±0.95)	1.75 (±0.5)	
2023										
Sampling site	Sampling date	Species	n	Weight (g)	Length (cm)	Found parasites	Parasites n=	Prevalence (%)	Abundance (± DP)	Intensity (± DP)
1 (Sado river)	01.02.2023	<i>Sparus aurata</i>	20	451.54 (±52.15)	30.65 (±1.15)	<i>Dactylogyrus</i> sp.	46	60	2.30 (±2.72)	3.83 (±1.9)
						<i>Sparocotyle chrysophrii</i>	39	50	1.95 (±2.39)	3.9 (±2.7)
2 (Ria de Alvor)	27.02.2023	<i>Sparus aurata</i>	28	486.16 (±4.84)	30.72 (±0.56)	<i>Furnestinia echeneis</i>	602	89.29	21.52 (±3.31)	24.08 (±2.109)
						<i>Diplectanum aequans</i>	3194	100	118.30 (±90.63)	118.30 (±9.388)
3 (Ria Formosa)	16.01.2023	<i>Dicentrarchus labrax</i>	27	264.42 (±22.68)	28.34 (±0.91)	<i>Lernanthropus kroyeri</i>	112	92.59	4.15 (±2.25)	4.48 (±1.9)
						<i>Caligus</i> sp.	13	40.74	0.48 (±0.64)	1.18 (±1.2)
4 (Ria de Aveiro)	11.04.2023	<i>Sparus aurata</i>	27	437.35 (±67.83)	30.53 (±2.29)	<i>Dactylogyrus</i> sp.	322	92.3	11.93 (±9.27)	12.39 (±1.46)
						<i>Dactylogyrus</i> sp.	573	100	30.16 (±29.35)	30.16 (±12.2)
4 (Ria de Aveiro)	11.04.2023	<i>Dicentrarchus labrax</i>	19	332.11 (±93.08)	31.51 (±2.01)	<i>Caligus</i> sp.	42	78.95	2.21 (±1.79)	2.8 (±1.6)
						<i>Praniza</i>	1	5.23	0.05 (±0.23)	1 (±0.1)
4 (Ria de Aveiro)	11.04.2023	<i>Sparus aurata</i>	12	342.81 (±60.32)	28.02 (±1.23)	<i>Dactylogyrus</i> sp.	76	100	6.34 (±4.64)	6.3 (±1.15)
						<i>Sparocotyle chrysophrii</i>	33	75	2.75 (±4.09)	3.67 (±1.13)
					<i>Caligus</i> sp.	2	16.67	0.17 (±0.39)	1 (±0.1)	
					<i>Praniza</i>	54	100	4.5 (±1.9)	4.5 (±2.8)	

Table 1. Identified parasites and statistical data associated with the infections.



LEGENDS:

- Figure 2 – *Trichodina* sp. (Protozoa: Ciliophora: Trichodinidae) in the gills
- Figure 3 – *Caligus* sp. (Copepoda: Caligidae)
- Figure 4 – *Praniza* larvae of a Gnathiid (Isopoda, Gnathiidae)
- Figure 5a – *Lernanthropus kroyeri*, parasitic Copepoda (Siphonostomatoidea, Lernanthropidae) attached horizontally to the gills
- Figure 5b – Adult female specimen of the parasitic copepod, *Lernanthropus kroyeri*
- Figure 6 – Adult female of the monogenean *Sparocotyle chrysophrii*. Presence of fertilised eggs within the uterus (*)
- Figure 7 – The monogenean *Dactylogyrus* sp.
- Figure 8 – *Diplectanum aequans* (Monogenea, Diplectanidae)

*The photos exhibited the morphological details of the parasites used to their identification

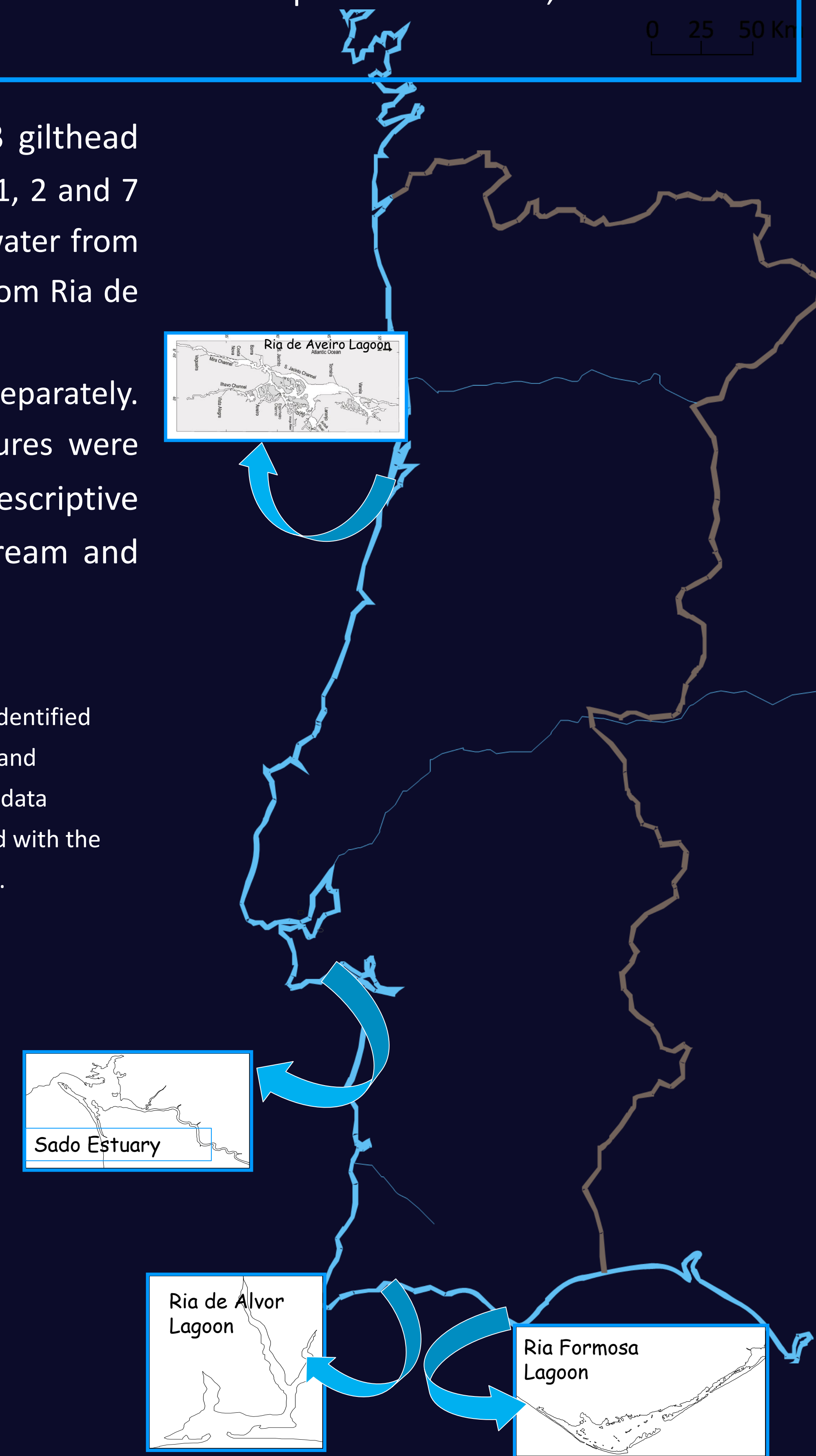


Fig. 1. Map of Portugal showing the fish farms location.

Three classes of parasites were found attached to the gills of the European seabass or gilthead seabream (Figures 2 to 9)[¶], namely the copepod crustaceans *Lernanthropus kroyeri*, *Caligus* sp. and the gnathiid isopod *Praniza*; the protozoan *Trichodina* sp. and the monogenean species *Diplectanum aequans*, *Dactylogyrus* sp., *Sparocotyle chrysophrii* and *Lamellodiscus* (syn. *Furnestinia*) *echeneis*. No nematodes were identified in the samples. In the 2023 samples, two new species of Monogenea were identified in both gilthead seabream and European seabass: *Dactylogyrus* sp. was identified in European seabass from one fish farm with production of both fish species. *Lamellodiscus* (syn. *Furnestinia*) *echeneis* is a new specimen identified in gilthead seabream.

There were no significant statistical differences between samples and sampling sites.

CONCLUSION: This paper addresses the concern that the presence of parasites in semi-intensive aquaculture systems can spread within and between semi-intensive fish farms and to wild fish by the seawater flow supplied by the tide. In these cases, infected fish farms should keep parasite density low at all times in order to reduce as much as possible the infection of neighbouring semi-intensive fish farms and wild fish. Three possible ways of infection of fish in these semi-intensive farms can be pointed out: a) infected fingerlings stocks were used to stock uninfected ponds; b) the characteristics of the production systems that make use of the tides for water renewal; c) the environmental conditions, namely the increase in water temperature that directly influences the parasite fauna of fishes. We must remain vigilant and monitor the evolution of the parasite infections.