

COMPARION OF DIGITAL COUNTING METHODS FOR KI67 STAINED MAST CELL TUMORS

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Introduction: Automated analysis of Ki67 expression is more and more commonly used in the daily routine diagnostics and prognostics of canine mast cell tumors. The purpose of this study was to compare different methods within the same tumour cohort and to find a practicable approach for the daily diagnostic routine

Materials and Methods: FFPE tissue from 78 archived canine mast cell tumors served as material. Slides were reviewed and separated in two groups (cutaneous and subcutaneous), cutaneous were graded according to Patnaik and Kiupel ^{1,2}. Routinely Giemsa and Ki67 stained slides were scanned and aligned using Visiopharm® for selection of tumour tissue. Ki67 positive nuclei were counted on the whole slide (output: % of total nuclei) using different methods followed by construction of a heat map. Positive cells were then counted in the hot spots according to published literature for manual and automated counting.

Table 1: Workflow of Ki67 counting











Alignement of Giemsa and Ki67 staining

 Automatically done Problems to solve: Preanalytical artefacts on slide, orientation of tissue on slide, scanning issues Example, Tumor H16-2495.4

Delineation of tumor tissue (Tissue detection App) Automatically done using the contrast of the metachromatic stain Problems to solve: variable metachromacy of mast cells Two Apps developed



Detection and exclusion of artefacts (Artefact App)

The algorithm took into consideration: Pixel differences in different colour channels, shape and additional parameters (size, neighboorhood etc).

All slides were run with the artefact Apps and afterwards visually controlled (4 eye principle) to find the most suitable App. Finally slides were categorized into two groups according to their staining intensity to find the best artefact for the slides.







Three different workflows were compared (a-c) followed by a Ki67 counting App(d-f): • a and d: Run of Ki67 App without artefact App

• b and e: artefact App and rund of Ki67 App without visual control • c and f : artefact App with visual control

a-c: dotted green line: region of interest (ROI) d-f: green: negative nuclei; red: positive nuclei

Heatmap and counting of cells in HOT SPOTs

A heatmap was created using label one (positive cells) within ROI 1 (tumor) with protected ROIS (excluded areas-artefacts). red: hotspot Inset a: 5 squares- method Inset b: total area of -method

A comparison was done between the total area of grids (free drawn object) and 5 squares (each equivalent to one grid area)

Results:

 Table 2
 Comparison of percentage of Ki67 positive cells over the whole tumor
 with/ without artefact exclusion (image a-f table 1)

Case Number	Percentage positive cells	Percentage positive cells	Percentage positive cells
	without artefact APP	with artefact APP	with artefact APP and
			visual control
	4 710072000	4.007636445	E 013564043
H16-2495.4_KI67	4.719673886	4.907030445	5.012564913
H16 2490.2_KI07	0.400959878	0.254409095	0.0754405
H10-2490.3_KI07	0.734974473	0.001563668	0.0754495
H16 2702 1 4:67	0.10494557	0.091303008	0.020303307
H16-2732.1_Ki67	1 842548845	0.22360/153	0.130982266
H17-0409 1 ki67	0 3312757	0.212212691	0.15707496
H17-0432.2 ki67	0.25214426	0.137133695	0.044545635
H17-0432.3 ki67	0.377695684	0.151895503	0.069002219
H17-0942.1 ki67	0.025447451	0.025518884	0.0234055
H17-1200.2 ki67	0.180124167	0.095449912	0.034690102
 H17-1449.1 ki67	0.310096098	0.26950695	0.195126673
 H17-1449.2_ki67	0.523352235	0.470359078	0.469064993
 H17-2069.1_ki67	0.235472662	0.136599706	0.062313837
	0.374152249	0.122226362	0.078542608
H17-2212.2_ki67	0.466036517	0.461945969	0.41687116
H17-2253.1_ki67	5.197121066	2.683214496	3.039451912
H17-2301.1_ki67	0.539245057	0.572694786	0.587976078
H17-2388.3_ki67	0.481625695	0.396056836	0.314260004
H17-2826.1_ki67	0.194733819	0.204907686	0.190944004
H18-0046.1_ki67	11.71983655	10.63384712	12.38511835
H18-0069.3_ki67	0.382260904	0.36078745	0.042902672
H18-0252.1_ki67	0.399382171	0.366578255	0.325128271
H18-0503.1_ki67	0.00252444	0.001381444	0.001731947
H18-1130.1_ki67	2.118735888	2.057569896	2.18589645
H18-1230.1_ki67	0.089495524	0.01225795	0.012620845
H18-1255.1_ki67	0.014139801	0.013990342	0.013668788
H18-1347.1_ki67	0.07592068	0.072770273	0.07117519
H18-1512.1_ki67	0.038383553	0.019383031	0.006016411
H18-1600.1_ki67	0.423797881	0.37422967	0.166895739
H18-1641.1_ki67	0.11322812	0.109609061	0.082586965
H18-1809.1_ki67	0.152047243	0.112712382	0.038230781
H18-1873.1_ki67	0.264707146	0.264199936	0.207298551
H18-2028.1_ki67	0.52320258	0.440534541	0.430993922
H18-2169.1_ki67	0.433715095	0.418809133	0.43887515
H18-2424.1_ki67	0.505408711	0.506950948	0.507360658
H18-2516.3_ki67	0.018345436	0.017845009	0.018475332
H18-2533.1_ki67	0.018656217	0.016224356	0.012931328
H18-2695.5_KI67	0.0231/5/77	0.021988896	0.016518253
H19-0407.1_KI67	0.019196216	0.018535323	0.011330849
H19-0438.5_Kl67	0.053974058	0.049086638	0.045823053
H19-0945 1 ki67	0.402010529	0.403/13255	0.403/1/826
H19-1335 1 ki67	0.24410591/	0.030343979	0.023895942
H19-1497 1 ki67	0.1707020705	0.142508080	0.150998409
H19-1649 1 ki67	0.032624703	0.022227143	0.021300237
H20-1046.4 ki67	0.461550198	0.415221419	0.347209181
H20-2061.1 ki67	0.117673285	0.710221419	0.013206603
H20-2525.2 ki67	1.254041772	1.272377243	1.019661414
H20-2670.1 ki67	1.362568626	1.2473674	1.177910743
H20-2699.3 ki67	0.28960201	0.235390662	0.067724206
H21-0807.1 ki67	0.831381087	0.76374788	0.822275405
H21-2935.1 kj67	1.578498243	1.58681442	1.592933097
H22-0073.1 ki67	0.496991891	0.49833887	0.191468993
H22-0537.1 ki67	4.046541642	3.634762506	3.99776847
H22-2000.1 ki67	2.653342522	1.860914748	2.053598314

Table 3: Statistical difference of three different workflows Bland-Altman plot for comparison of Percentage of Ki67 positive cells

without artefact app compared to with artefact App without visual control.

Results: The workflows allowed standardised collection and comparison of the Ki67 index as well as selection of Hotspots. Alignment of Giemsa and Ki67 stained slides was useful for the detection of tumour tissue. Additional visual exclusion of artefacts did not have a significant effect on the counting of positive cells within the whole tumour after application of the artefact APP (Bland Altman plot 1 and



2). Visual control of the hot spots appeared necessary to avoid that melanophages and blood vessels might lead to false positives. The selection of the method (square versus area) in the workflow grid influences the outcome of cases above the threshold.

Table 4: Comparison of cases above the threshold in different Ki67 positive counting methods



Table 5: Distribution of tumor grades according to Patnaik and Kiupel among cutaneous tumors

Patnaik Grading

Kiupel Grading

Conclusions:

This study underlines the need of a standardized approach for the automated Ki67 counting in mast cell tumours among pathologists, which allows comparison of different methods^{3,4} with fewer resources. Examination of a larger cohort with clinical data is now needed to compare the different methods. Techniques published for the microscopic evaluation pose some difficulties when transferred to the automatic counting. In addition, transparent communication and close collaboration with the laboratory is needed to minimize preanalytical issues and thereby saves pathologists time and costs.



Literature:

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